

ScienceDirect



Restoring vision at the fovea Juliette E McGregor



In humans high quality, high acuity visual experience is mediated by the fovea, a tiny, specialized patch of retina containing the locus of fixation. Despite this, vision restoration strategies are typically developed in animal models without a fovea. While electrical prostheses have been approved by regulators, as yet they have failed to restore high quality, high acuity vision in patients. Approaches under pre-clinical development include regenerative cell therapies, optogenetics and chemical photosensitizers. All retinal vision restoration therapies require reactivation of inner retina that has lost photoreceptor input and that the restored signals can be interpreted at a behavioural level. A greater emphasis on tackling these challenges at the fovea may accelerate progress toward high quality vision restoration.

Address

Center for Visual Science, University of Rochester, 601 Crittenden Blvd, Rochester, NY, USA

Corresponding author: McGregor, Juliette E (jmcgrego@ur.rochester.edu)

Current Opinion in Behavioral Sciences 2019, 30:210-216

This review comes from a themed issue on **Visual perception** Edited by **Hannah Smithson** and **John S Werner**

https://doi.org/10.1016/j.cobeha.2019.10.003

2352-1546/© 2019 Published by Elsevier Ltd.

Introduction

The human fovea is specialized for high acuity vision, contains the locus of fixation and mediates much of our conscious visual experience, yet it represents only a tiny patch of retina subtending less than the size of your thumbnail at arm's length. Age related macular degeneration (AMD), the leading cause of blindness in the developed world, begins with damage to the photoreceptors in this region and the loss of central vision. Vision restoration approaches which include the fovea have been shown to be more effective [1] perhaps because this tiny patch of retina provides input to a highly expanded representation within the visual brain [2]. Despite these compelling reasons to study it, much of the preclinical development is performed on animals without a fovea. In this review we consider how the vision restoration techniques currently being explored by the scientific

community interact with the unique structure and physiology of the fovea and evaluate the prospects for restoring high quality foveal vision. We examine how functional outcomes at the fovea can be evaluated at preclinical and clinical stages and highlight key unanswered questions.

The structure of the fovea

The fovea has unique anatomical and physiological specializations [3] presenting both special challenges, and certain advantages for vision restoration. The fovea is avascular and foveal cones are slender and densely packed relative to their peripheral counterparts. This creates a high metabolic burden, which may make the fovea vulnerable to degeneration [4]. The inner retina, containing the neurons that perform the initial stages of visual processing are also specialised in this region. High acuity in the fovea is mediated by an expansion of the midget retinal ganglion cell (RGCs) class. Retinal physiology is reviewed in detail elsewhere [5], but here we note that unlike in the periphery where signals from large numbers of photoreceptors are pooled, each foveal midget RGC has a 'private line' through dedicated bipolar cell to an individual cone. This means the density of retinal ganglion cells at the fovea, and the area of visual cortex devoted to processing foveal signals, is increased relative to the periphery. While colour perception is a feature of foveal vision, from a restoration standpoint this is likely to be less important to quality of life than gains in acuity. From an evolutionary standpoint colour vision was a late addition to the fovea and that high acuity spatial vision brought about by high cone density and the specialized midget circuitry was the primary aim of the structure [5].

Directly above the region of high cone density, the inner retinal layers are thinned, creating perhaps the most distinctive anatomical feature of the fovea, the 'foveal pit'. The 200 µm floor of the pit, referred to as the 'foveola' is particularly thin and delicate. The foveal pit is formed during development where foveal RGCs are laterally displaced by hundreds of microns relative to the cones they are connected to. The radially displaced retinal ganglion cell somas pile up in a ring on the margins of the pit as shown in Figure 1. Recent experiments have shown that despite the huge distances moved by the ganglion cells relative to the cones they are connected to, the local spatial arrangement of retinal ganglion cell somas within the ring closely matches the relative arrangement of receptive field locations [6[•]]. This becomes important if we seek to bypass degenerated foveal cones and restore vision by directly stimulating the displaced ganglion cells. The thin inner limiting



Illustration of vision restoration strategies at the fovea. Retinal ganglion cells are displaced from the foveal photoreceptors they serve (red line). This results in the formation of an RGC ring (green) around the foveal pit, the floor of which is referred to as the foveola. A selection of vision restoration strategies are shown: subretinal approaches (electrical prostheses, photoreceptor precursor transplantation), intravitreal (chemical photosensitizers and inner retina optogenetic) and the epi-retinal prosthesis. Abbreviations: GCL - Ganglion cell layer, INL - Inner nuclear layer, ONL - Outer nuclear layer, PRL – Photoreceptor layer.

membrane at the foveola is advantageous for restoration approaches that rely on intravitreal injections, allowing increased penetration of therapeutics into the retinal tissue at this location [7].

The current state of play

The only vision restoration therapy currently approved by regulators in the EU and the USA is the epiretinal Argus II electrical prosthesis. An electrode array is surgically implanted on the surface of the retina centred on the fovea but extending well beyond its margins. A head mounted camera converts the scene into patterns of electrical stimulation that are applied to RGCs [8]. One drawback of such prostheses is the sparsity of the stimulating electrodes and at present it is not possible to restore vision at acuities close to that of normal fovea [9]. Nevertheless, recipients of this device may see and interact with low spatial frequency, high contrast stimuli like a white ball rolling on a black table and show improvements in activities of daily living [10,11]. In Europe two sub retinal implants, the Alpha IMS and Alpha AMS (wireless version) are approved; in theory these offer an eightfold increase in resolution, however in practice, the achievable acuity is similar to the Argus II [12,13,14[•],15]. A large range of electrical retinal prostheses are under development [16] and efforts are ongoing to develop higher resolution photovoltaic implants that are better able to confine the electric field and achieve closer contact with the target cells [17,18]. Higher acuity has been achieved in rodents [19] but these improvements have yet to be translated into primates and humans. Cortical magnification of the foveal region means that chip implantation directly in cortex may be a promising future direction [11,20] although magnification factor does vary significantly between individuals [21].

Hopes of high-quality vision restoration have long lain in regenerative therapies which offer the possibility of regrowing replacements for degenerated cones from transplanted stem cell derived photoreceptor precursor cells. This approach is currently in its early stages, with researchers actively working to establish photoreceptor precursor survival, maturation, synaptogenesis and integration with the host following cell delivery into the subretinal space [22–24]. Potential synaptic interactions between transplanted retinal sheets and the host have been observed in rodent models of retinal degeneration [25,26] and primate [27]. The fovea presents a challenge for transplantation because of tissue delicacy and the displacement of retinal ganglion cells relative to the native photoreceptors that drive them. During development RGC-photoreceptor connections are made before the lateral displacement occurs. Photoreceptors transplanted subretinally into the adult foveola would need to extend axons hundreds of microns laterally to connect with their target RGCs in the foveal ring. An alternative approach would be to deliver cone precursors to the subretinal space directly under the ring of foveal RGCs to maximize the possibility of interaction. The impact of the degenerating environment on transplanted cells, particularly on cone function is still poorly understood [23,28,29]. In vivo retinal imaging of fluorescently labelled photoreceptor precursor cells has allowed longitudinal monitoring of transplant survival and remodelling in the intact non-human primate [30].

Instead of re-growing new photoreceptors, an alternative vision restoration approach involves repurposing the remaining architecture. Optogenetic vision restoration involves inserting light-sensitive ion channels directly into inner retinal neurons, making them intrinsically light sensitive and thus overcoming the loss of photoreceptor input. This is achieved by infecting inner retina with a virus that codes for the optogenetic actuator which is then synthesised by the infected cells themselves. Using custom promoters it is possible to target-specific cell classes like RGCs [31^{••}] or bipolar cells [32,33]. The cell class chosen may affect the gain and the type of restored responses. The type of bipolar cells to which an RGC is connected, dictates whether it is an 'On center' RGC, increasing its activity in response to a light increment, or an 'off center' RGC, increasing its activity in response to a light decrement. Inserting channels into bipolar cells could potentially restore both 'on' and 'off' responses, whereas inserting the same channel directly into RGCs would turn every RGC into an 'on center' type. The psychophysical consequences of these choices are largely unexplored. Optogenetic actuators are currently relatively light insensitive as they are based on microbial opsins, but higher sensitivity actuators based on mammalian opsins and custom engineered G-protein coupled receptors are becoming available [34,35^{••},36,37].

Optogenetic vision restoration requires only a single injection and offers potentially high acuity, as rather than a sparse array of electrodes, every cell can be light sensitive. The drawbacks of using the foveal ring as a light sensor is that many cells are piled up on top of each other and the positions of the ganglion cells are dilated into a ring relative to the original positions of their foveal receptive fields [6[•]]. This may result in a perceived warping of the visual stimulus shape and size and a 'blind spot' in the foveal center. It may be possible to pre-distort the stimulus to compensate, or patients may adapt to this form of vision as they do to vision loss [38,39]. Head mounted optogenetic visual stimulators [40] are being developed, including versions with encoders that present the anticipated RGC firing pattern a scene would evoke, rather than the scene itself [41]. Optogenetic vision restoration has been demonstrated ex-vivo in foveal tissue [31^{••},42] and recently in the primate *in vivo* [43]. The perceptual and behavioural consequences of this unusual form of vision, remain unexplored.

As some photoreceptor inner segments are functionally preserved in degenerating retina [44], it may be possible to deliver optogenetic gene therapy directly to these cells, maintaining the spatial layout of the intact retina. This has been demonstrated *ex-vivo* in human retina [44] and offers the most straightforward route to naturalistic vision with the lightest requirement for adult plasticity. Recent efforts have focussed on the development of vectorpromoter combinations that can produce expression in foveal cones via intravitreal rather than subretinal injection, avoiding detachment of the delicate foveola [45]. Viral transduction across the RPE may also be possible using ultrasonic techniques [46].

It is also possible to make the inner retina light sensitive using azobenzene-based photoswitches such as Benaq [47[•]]. This molecule can be delivered intravitreally and in mice preferentially targets retinal ganglion cells that have undergone physiological changes due to the loss of photoreceptor input, selectively restoring 'off responses' [48]. A major difference from optogenetic therapy, is that chemical photosensitizers are cleared from the retina in a month, necessitating repeated injections. This therapy has not yet been tested in a foveate animal and therefore whether sensitization is confined to the RGC ring or works pan-retinally is unknown.

All retinal vision restoration therapies rely on the functional preservation of retinal ganglion cells despite loss of photoreceptor input. Unfortunately, RGCs can undergo functional changes following photoreceptor loss including hyperactivity and remodelling [49,50–53] which may reduce the quality of the restored vision in the human [54]. It has been suggested that retinoic acid diffusing from RPE at the site of photoreceptor degeneration, into the ganglion cell layer, is the signal which triggers hyperpermeability of RGCs and this leads to hyperactivity [55[•]]. If this hypothesis is true, foveal RGCs may be less vulnerable to locally diffusing retinoic in the initial stages of macular degeneration than mouse models would suggest because foveal RGCs are displaced relative to region of degenerating photoreceptors. Regardless of the trigger, there is evidence that gap junction blockers like meclofenamic acid, can reduce hyperactivity and enhance visual performance in mice [56] but this solution has yet to be tested in large animal models or humans. As gap junctions create an electrical syncytium in the retina, disrupting these channels prevents waves of spontaneous activity spreading through the tissue.

Similar questions surround preventing or reversing pathological changes taking place to the retinal blood supply following photoreceptor degeneration [57,58]. Progressive changes in blood flow that have developed over many years of disease may result in a limited oxygen supply being available to 'restored' retinal ganglion cells, whose altered physiology may be more energy intensive. There are many unanswered questions surrounding reintroducing function to a tissue that has been deprived of normal activity patterns for days, months or years. Functional and structural changes may be easier to combat if detected early clinical trials for vision restoration are typically performed in challenging end stage cases where little information can be gleaned. Little is currently known as to how to preserve or treat the retina to provide the biological infrastructure needed to achieve high quality vision restoration.

Toward high quality vision restoration at the fovea

To overcome the challenges of vision restoration at the fovea and accelerate progress toward successful phase III clinical trials, preclinical studies are necessary. Unfortunately, there is a paucity of animal models with human like-foveal specializations and within that group even fewer models of retinal degeneration. The mouse, the most commonly used laboratory animal [35^{••},59,60] has several genetic models of degeneration but no fovea-like specializations. In canine species several genetic degenerations have been discovered [61] and there is an 'area centralis' containing a small bouquet of cones at a density similar to that of the primate [62], however there is no foveal excavation or ganglion cell specialization. The ground squirrel has a cone dominated retina, with an 'area centralis' a larger area of densely packed cones [63] and an increased ganglion cell density but no foveal excavation or avascular zone [64]. This species has been used to explore circuit repair [49] due to the retinal changes that take place during hibernation. Other cone-dominant species continue to be explored as potential animal models for retinal disease [65,66].

Primates have the most similar foveal structure to humans, with macaque monkeys being the pre-clinical species of choice. Old world primates develop drusen [67] and an inherited form of achromatopsia was recently discovered in the rhesus macaque [68^{••}], however at present, acute and inducible forms of vision loss, such as laser lesions [69] and cobalt chloride lesions [27] are more common. The marmoset, which is smaller and easier to breed has potential as another foveate model species [70,71]. Transgenic degeneration models in primate may soon be possible [72,73].

To evaluate the success of vision restoration therapies at the pre-clinical stage *in vivo*, it's possible to express the fluorescent calcium indicator GCaMP in foveal RGCs and use adaptive optics ophthalmoscopy to non-invasively read out cellular activity [74]. This method has demonstrated the restoration of pattern vision at the retinal level in the macaque [43]. In the future it may be possible to monitor restored activity using intrinsic signals [75,76] or voltage indicators [77]. The cortical magnification of foveal projections to the brain may allow recording of visually evoked potentials from primate fovea. This would allow us to assess if restoration at the retinal level translates to stimulation in higher nuclei. Ultimately psychophysics will provide the gold standard to evaluate how the brain is able to interpret the restored visual cues.

Currently all prospective vision restoration therapies can only 'restore' an impoverished form of vision, nevertheless patients are receiving electrical retinal prostheses. and optogenetic therapies are entering clinical trials. Amid the complexities of the diseased retina it can be difficult to distinguish cause and effect when studying vision restoration and testing is not yet fully standardized. In the clinic, Humphery visual field testing (perimetry) is common and at the fovea, higher density 'microperimetry' is used, however this is still very coarse compared to the grain of the photoreceptor mosaic. Adaptive optics ophthalmoscopy has been utilized to perform visual testing at high resolution [78] and this could be applied map foveal function before and after intervention. Visual field testing can be a problem for patients with foveal impairment if they are not able to fixate. In such cases tracking eye movements and the recovery of a stable fixation locus could be used to evaluate the success of an intervention.

Visual acuity is an appropriate metric to assess foveal vision and is pertinent to quality of life as the fovea mediates detailed spatial vision needed for reading, recognising faces and interpreting facial expressions. While valuable, acuity alone does not evaluate the richness of visual experience or usability of the restored function. Behavioural tests such as the multi-luminance mobility test have been developed [79] and for the fovea tests of contrast sensitivity [80], localization, shape and object perception are also informative. In ultra-low vision patient populations receiving the first generation of restorative therapies, acuity may not be measurable. Researchers have been working with patients to develop quality of life and visual function questionnaires capable of capturing a fuller picture of the impact of a therapy [81-83].

Lastly it should be noted that all efforts to restore vision at the retinal level are constrained by levels of plasticity in the adult brain which must interpret the restored signals. This is particularly relevant if the form of restored vision is very different from natural vision. Evidence from patients who have received retinal prostheses suggests that low resolution 'pixelated' scenes can be interpreted. Higher resolution forms of restored vision relying on direct activation of the RGC ring will require distortion of the stimulus or some degree of plasticity to achieve high acuity in practice. Visual rehabilitation training [84] is likely to be necessary for any form of vision restoration and whilst there is evidence of plasticity in the adult, it is limited [39]. In canine and murine pre-clinical studies there is evidence that younger animals show a greater visual rescue following retinal gene therapy than older animals [85,86]. A similar age dependency has been reported in clinical trials [87] suggesting that once degeneration is too advanced rescue may be more limited. The increasing deployment of vision restoration therapies in preclinical models and patients may afford an opportunity to learn more about the fundamentals of both retinal rescue and visual plasticity of downstream nuclei. Primate psychophysics may help us to assess the bounds of usability of these forms of restored vision before they enter expensive clinical trials.

Conclusion

While current interventions cannot yet provide the highquality vision that the term 'vision restoration' implies, a range of promising therapies are under development. Devising strategies which are cognisant of the unique feaures of the fovea may accelerate progress toward this goal. Access to new primate models of retinal degeneration and methods of evaluating function *in vivo* may make pre-clinical studies of foveal vision restoration more readily achievable, improving the quality of therapies entering clinical trials.

Conflict of interest statement

Nothing declared.

Funding

This work was supported by the National Eye Institute [NIH RO1 EY021166, NIH UO1 EY025497]. The funding source had no role in decisions relating to this manuscript.

Acknowledgements

We acknowledge William Merigan for feedback on the manuscript and William Merigan and David Williams for helpful discussions.

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- · of special interest
- •• of outstanding interest
- Stingl K, Bartz-Schmidt K-U, Gekeler F, Kusnyerik A, Sachs H, Zrenner E: Functional outcome in subretinal electronic implants depends on foveal eccentricitysubretinal electronic implants. Invest Ophthalmol Vis Sci 2013, 54:7658-7665.
- Tootell R, Switkes E, Silverman M, Hamilton S: Functional anatomy of macaque striate cortex. II. Retinotopic organization. J Neurosci 1988, 8:1531.
- Hendrickson A: Organization of the adult primate fovea. In Macular Degeneration. Edited by Penfold PL, Provis JM. Berlin Heidelberg: Springer; 2005:1-23.
- Provis JM, Penfold PL, Cornish EE, Sandercoe TM, Madigan MC: Anatomy and development of the macula: specialisation and the vulnerability to macular degeneration. *Clin Exp Optom* 2005, 88:269-281.
- Thoreson WB, Dacey DM: Diverse cell types, circuits, and mechanisms for color vision in the vertebrate retina. *Physiol Rev* 2019, 99:1527-1573.

- McGregor JE, Yin L, Yang Q, Godat T, Huynh KT, Zhang J,
 Williams DR, Merigan WH: Functional architecture of the
- Williams DR, Merigan WH: Functional architecture of the foveola revealed in the living primate. PLoS One 2018, 13: e0207102

This study shows that there is an orderly mapping between the positions of retinal ganglion cells in the foveal ring and the receptive fields they serve. This mapping implies that locally, small visual stimuli presented directly to light sensitive foveal ganglion cells could evoke a similar spatial pattern of activity to natural vision. Globally however, spatial distortions to stimuli are expected at the fovea.

- 7. Dalkara D, Kolstad KD, Caporale N, Visel M, Klimczak RR, Schaffer DV, Flannery JG: Inner limiting membrane barriers to AAV-mediated retinal transduction from the vitreous. *Mol Ther* 2009, **17**:2096-2102.
- 8. Luo YH-L, da Cruz L: The Argus® II retinal prosthesis system. *Prog Retin Eye Res* 2016, **50**:89-107.
- Damle S, Lo Y-H, Freeman WR: High visual acuity retinal prosthesis: understanding limitations and advancements toward functional prosthetic vision. *Retina* 2017, 37.
- da Cruz L, Dorn JD, Humayun MS, Dagnelie G, Handa J, Barale P-O, Sahel J-A, Stanga PE, Hafezi F, Safran AB et al.: Five-year safety and performance results from the Argus II retinal prosthesis system clinical trial. Ophthalmology 2016, 123:2248-2254.
- 11. Stronks HC, Dagnelie G: The functional performance of the Argus II retinal prosthesis. Expert Rev Med Devices 2014, 11:23-30.
- 12. Edwards TL, Cottriall CL, Xue K, Simunovic MP, Ramsden JD, Zrenner E, MacLaren RE: Assessment of the electronic retinal implant alpha AMS in restoring vision to blind patients with end-stage retinitis pigmentosa. *Ophthalmology* 2018, **125**: 432-443.
- 13. Mills JO, Jalil A, Stanga PE: Electronic retinal implants and artificial vision: journey and present. *Eye* 2017, **31**:1383.
- Stingl K, Schippert R, Bartz-Schmidt KU, Besch D, Cottriall CL,
 Edwards TL, Gekeler F, Greppmaier U, Kiel K, Koitschev A et al.: Interim results of a multicenter trial with the new electronic subretinal implant alpha AMS in 15 patients blind from inherited retinal decenerations. Front Neurosci 2017. 11:445

inherited retinal degenerations. Front Neurosci 2017, 11:445 The authors use a battery of clinical tests to evaluate the functional outcomes of the most advanced subretinal implant approved for use in humans, the Alpha AMS.

- Stingl K, Bartz-Schmidt KU, Besch D, Chee CK, Cottriall CL, Gekeler F, Groppe M, Jackson TL, MacLaren RE, Koitschev A et al.: Subretinal visual implant alpha IMS – clinical trial interim report. Sight Restor Prosthet Optogenetics Gene Ther 2015, 111:149-160.
- Bloch E, Luo Y, da Cruz L: Advances in retinal prosthesis systems. Ther Adv Ophthalmol 2019, 11:2515841418817501.
- Boinagrov D, Pangratz-Fuehrer S, Goetz G, Palanker D: Selectivity of direct and network-mediated stimulation of the retinal ganglion cells with epi-, sub- and intraretinal electrodes. J Neural Eng 2014, 11:026008.
- Flores T, Lei X, Huang T, Lorach H, Dalal R, Galambos L, Kamins T, Mathieson K, Palanker D: Optimization of pillar electrodes in subretinal prosthesis for enhanced proximity to target neurons. J Neural Eng 2018, 15:036011.
- Lorach H, Goetz G, Smith R, Lei X, Mandel Y, Kamins T, Mathieson K, Huie P, Harris J, Sher A et al.: Photovoltaic restoration of sight with high visual acuity. Nat Med 2015, 21:476.
- 20. Niketeghad S, Pouratian N: Brain machine interfaces for vision restoration: the current state of cortical visual prosthetics. *Neurotherapeutics* 2018, **16**:134-143.
- 21. Harvey BM, Dumoulin SO: The relationship between cortical magnification factor and population receptive field size in human visual cortex: constancies in cortical architecture. *J* Neurosci 2011, **31**:13604.
- 22. Gamm DM, Wong R, the AGI Workshop Panelists: Report on the national eye institute audacious goals initiative:

photoreceptor regeneration and integration workshop. Transl lis Sci Technol 2015, 4:2

- 23. Gasparini SJ, Llonch S, Borsch O, Ader M: Transplantation of photoreceptors into the degenerative retina: current state and future perspectives. Prog Retin Eye Res 2018, 69:1-37 http://dx. doi.org/10.1016/j.preteyeres.2018.11.001.
- 24. Waldron PV. Di Marco F. Kruczek K. Ribeiro J. Graca AB. Hippert C, Aghaizu ND, Kalargyrou AA, Barber AC, Grimaldi G et al.: Transplanted donor- or stem cell-derived cone photoreceptors can both integrate and undergo material transfer in an environment-dependent manner. Stem Cell Rep 2018. 10:406-421.
- 25. Foik AT, Lean GA, Scholl LR, McLelland BT, Mathur A, Aramant RB, Seiler MJ, Lyon DC: Detailed visual cortical responses generated by retinal sheet transplants in rats with severe retinal degeneration. J Neurosci 2018, 38:10709.
- Iraha S, Tu H-Y, Yamasaki S, Kagawa T, Goto M, Takahashi R, Watanabe T, Sugita S, Yonemura S, Sunagawa GA *et al.*: Establishment of immunodeficient retinal degeneration model mice and functional maturation of human ESC-derived retinal sheets after transplantation. Stem Cell Rep 2018, 10:1059-1074.
- 27. Shirai H, Mandai M, Matsushita K, Kuwahara A, Yonemura S, Nakano T, Assawachananont J, Kimura T, Saito K, Terasaki H et al.: Transplantation of human embryonic stem cell-derived retinal tissue in two primate models of retinal degeneration. Proc Natl Acad Sci U S A 2016, 113:E81.
- 28. Pearson RA, Hippert C, Graca AB, Barber AC: Photoreceptor replacement therapy: challenges presented by the diseased recipient retinal environment. Vis Neurosci 2014, 31:333-344.
- Tsai ELS, Ortin-Martinez A, Gurdita A, Comanita L, Yan N, Smiley S, Delplace V, Shoichet MS, Nickerson PEB, Wallace VA: 29 Modeling of photoreceptor donor-host interaction following transplantation reveals a role for Crx, Müller glia, and Rho/ ROCK signaling in neurite outgrowth. Stem Cells 2019, 37.
- 30. McGregor JE, Phillips MJ, Walters S, Zhang J, Strazzeri J DiLoreto D, Walker A, Fischer WS, Yang Q, DiVincenti L: Noninvasive retinal imaging of fluorescent hESC-derived photoreceptor precursors in the living primate. Invest Ophthalmol Vis Sci 2017, 58:4576.
- Chaffiol A, Caplette R, Jaillard C, Brazhnikova E, Desrosiers M,
 Dubus E, Duhamel L, Macé E, Marre O, Benoit P *et al.*: A new promoter allows optogenetic vision restoration with enhanced sensitivity in macaque retina. Mol Ther 2017, 25:2546-2560

This study addresses the challenges of translating optogenetic therapies into the primate, including consideration of immune response, the development of a ganglion cell-specific promotor and goes on to test the therapy ex-vivo in macaque fovea.

- Cronin T, Vandenberghe LH, Hantz P, Juttner J, Reimann A, Kacsó Á, Huckfeldt RM, Busskamp V, Kohler H, Lagali PS et al.: 32. Efficient transduction and optogenetic stimulation of retinal bipolar cells by a synthetic adeno-associated virus capsid and promoter. EMBO Mol Med 2014, 6:1175.
- Lu Q, Ganjawala TH, Ivanova E, Cheng JG, Troilo D, Pan Z-H: AAV-mediated transduction and targeting of retinal bipolar cells with improved mGu R6 promoters in rodents and primates. Gene Ther 2016. 23:680.
- 34. Baker CK, Flannery JG: Innovative optogenetic strategies for vision restoration. Front Cell Neurosci 2018, 12:316.
- 35.
- Berry MH, Holt A, Levitz J, Broichhagen J, Gaub BM, Visel M, Stanley C, Aghi K, Kim YJ, Cao K *et al.*: **Restoration of patterned** vision with an engineered photoactivatable G protein-coupled receptor. Nat Commun 2017, 8:1862

Development of a high sensitivity optogenetic actuator based on the mammalian GPCR that can operate at ambient light levels and adapt. Patterned vision is demonstrated behaviourally in the mouse.

- Gaub BM, Berry MH, Holt AE, Isacoff EY, Flannery JG: 36. Optogenetic vision restoration using rhodopsin for enhanced sensitivity. Mol Ther 2015, 23:1562-1571.
- 37. van Wyk M, Pielecka-Fortuna J, Löwel S, Kleinlogel S: Restoring the ON switch in blind retinas: opto-mGluR6, a next-

generation, cell-tailored optogenetic tool. PLoS Biol 2015, 13: e1002143

- 38. Legge GE, Chung STL: Low vision and plasticity: implications for rehabilitation. Annu Rev Vis Sci 2016, 2:321-343.
- 39. Beveler M. Rokem A. Boynton GM. Fine I: Learning to see again: biological constraints on cortical plasticity and the implications for sight restoration technologies. J Neural Eng 2017. 14:051003.
- 40. Soltan A, Barrett JM, Maaskant P, Armstrong N, Al-Atabany W, Chaudet L, Neil M, Sernagor E, Degenaar P: A head mounted device stimulator for optogenetic retinal prosthesis. J Neural Eng 2018, 15:065002.
- 41. Nirenberg S, Pandarinath C: Retinal prosthetic strategy with the capacity to restore normal vision. Proc Natl Acad Sci U S A 2012, 109:15012.
- Sengupta A, Chaffiol A, Macé E, Caplette R, Desrosiers M, Lampič M, Forster V, Marre O, Lin JY, Sahel J et al.: Red-shifted channelrhodopsin stimulation restores light responses in blind mice, macaque retina, and human retina. EMBO Mol Med 2016. 8:1248.
- 43. McGregor JE, Godat T, Parkins K, Strazzeri J, Williams DR, Merigan WH: Channelrhodopsin mediated retinal ganglion cell responses in the living macaque. Invest Ophthalmol Vis Sci 2018, 59:2589.
- 44. Busskamp V, Duebel J, Balya D, Fradot M, Viney TJ, Siegert S, Groner AC, Cabuy E, Forster V, Seeliger M *et al.*: Genetic reactivation of cone photoreceptors restores visual responses in retinitis pigmentosa. Science 2010, 329:413.
- 45. Khabou H, Garita-Hernandez M, Chaffiol A, Reichman S, Jaillard C, Brazhnikova E, Bertin S, Forster V, Desrosiers M, Winckler C et al.: Noninvasive gene delivery to foveal cones for vision restoration. JCI Insight 2018, 3:e96029.
- Wang S, Kugelman T, Buch A, Herman M, Han Y, Karakatsani ME, Hussaini SA, Duff K, Konofagou EE: Non-invasive, focused ultrasound-facilitated gene delivery for optogenetics. Sci Rep 2017, 7:39955.
- Tochitsky I, Polosukhina A, Degtyar VE, Gallerani N, Smith CM,
 Friedman A, Van Gelder RN, Trauner D, Kaufer D, Kramer RH: Restoring visual function to blind mice with a photoswitch that exploits electrophysiological remodeling of retinal ganglion cells. Neuron 2014, 81:800-813

Demonstration that a chemical photosensitizer can restore light sensitivity in rodents for up to a month. This form of vision restoration is interesting as it only photosensitizes RGCs that have lost their photoreceptor input.

- 48. Tochitsky I, Helft Z, Meseguer V, Fletcher RB, Vessey KA, Telias M, Denlinger B, Malis J, Fletcher EL, Kramer RH: How azobenzene photoswitches restore visual responses to the blind retina. Neuron 2016, 92:100-113.
- 49. Beier C, Hovhannisyan A, Weiser S, Kung J, Lee S, Lee DY, Huie P, Dalal R, Palanker D, Sher A: Deafferented adult rod bipolar cells create new synapses with photoreceptors to restore vision. J Neurosci 2017, 37:4635.
- 50. Jones BW, Watt CB, Marc RE: Retinal remodelling. Clin Exp Optom 2005, 88:282-291.
- 51. Marc RE, Jones BW, Watt CB, Strettoi E: Neural remodeling in retinal degeneration. Prog Retin Eye Res 2003, 22:607-655.
- 52. Sekirnjak C, Jepson LH, Hottowy P, Sher A, Dabrowski W, Litke AM, Chichilnisky EJ: Changes in physiological properties of rat ganglion cells during retinal degeneration. J Neurophysiol 2011. 105:2560-2571.
- Stasheff SF: Emergence of sustained spontaneous 53. hyperactivity and temporary preservation of off responses in ganglion cells of the retinal degeneration (rd1) mouse. J Neurophysiol 2008, 99:1408-1421.
- 54. Stasheff SF: Clinical impact of spontaneous hyperactivity in degenerating retinas: significance for diagnosis, symptoms, and treatment. Front Cell Neurosci 2018, 12:298.

- 55. Telias M, Denlinger B, Helft Z, Thornton C, Beckwith-Cohen B,
- Kramer RH: Retinoic acid induces hyperactivity, and blocking its receptor unmasks light responses and augments vision in retinal degeneration. *Neuron* 2019, 102:574-586 http://dx.doi. org/10.1016/j.neuron.2019.02.015

Identification of retinoic acid as a diffusible signal triggering RGC hyperactivity in the rodent following photoreceptor degeneration including strategies to block the signal. If hyperactivity is also present in degenerated primate fovea, it will likely degrade the quality of restored vision.

- Barrett JM, Hilgen G, Sernagor E: Dampening spontaneous activity improves the light sensitivity and spatial acuity of optogenetic retinal prosthetic responses. Sci Rep 2016, 6:33565.
- Milam AH, Li Z-Y, Fariss RN: Histopathology of the human retina in retinitis pigmentosa. Prog Retin Eye Res 1998, 17:175-205.
- Hanna J, Yücel YH, Zhou X, Mathieu E, Paczka-Giorgi LA, Gupta N: Progressive loss of retinal blood vessels in a live model of retinitis pigmentosa. Can J Ophthalmol 2018, 53:391-401.
- 59. Lorach H, Goetz G, Mandel Y, Lei X, Kamins TI, Mathieson K, Huie P, Dalal R, Harris JS, Palanker D: Performance of photovoltaic arrays in-vivo and characteristics of prosthetic vision in animals with retinal degeneration. Sight Restor Prosthet Optogenetics Gene Ther 2015, 111:142-148.
- Lu Q, Ganjawala TH, Hattar S, Abrams GW, Pan Z-H: A robust optomotor assay for assessing the efficacy of optogenetic tools for vision restoration optomotor assay for optogenetic vision restoration. Invest Ophthalmol Vis Sci 2018, 59:1288-1294.
- Petersen-Jones SM, Komáromy AM: Dog models for blinding inherited retinal dystrophies. Hum Gene Ther Clin Dev 2014, 26:15-26.
- Beltran WA, Cideciyan AV, Guziewicz KE, Iwabe S, Swider M, Scott EM, Savina SV, Ruthel G, Stefano F, Zhang L *et al.*: Canine retina has a primate fovea-like bouquet of cone photoreceptors which is affected by inherited macular degenerations. *PLoS One* 2014, 9:e90390.
- Sajdak B, Sulai YN, Langlo CS, Luna G, Fisher SK, Merriman DK, Dubra A: Noninvasive imaging of the thirteen-lined ground squirrel photoreceptor mosaic. Vis Neurosci 2016, 33:e003.
- 64. Long KO, Fisher SK: The distributions of photoreceptors and ganglion cells in the California ground squirrel, *Spermophilus beecheyi*. *J Comp Neurol* 1983, **221**:329-340.
- 65. Verra DM, Sajdak BS, Merriman DK, Hicks D: Diurnal rodents as pertinent animal models of human retinal physiology and pathology. *Prog Retin Eye Res* 2019 http://dx.doi.org/10.1016/j. preteyeres.2019.100776. in press.
- 66. Sajdak BS, Salmon AE, Cava JA, Allen KP, Freling S, Ramamirtham R, Norton TT, Roorda A, Carroll J: Noninvasive imaging of the tree shrew eye: wavefront analysis and retinal imaging with correlative histology. *Exp Eye Res* 2019, 185:107683.
- Fletcher EL, Jobling AI, Greferath U, Mills SA, Waugh M, Ho T, de longh RU, Phipps JA, Vessey KA: Studying age-related macular degeneration using animal models. Optom Vis Sci Off Publ Am Acad Optom 2014, 91:878-886.
- Moshiri A, Chen R, Kim S, Harris RA, Li Y, Raveendran M, Davis S,
 Liang Q, Pomerantz O, Wang J et al.: A nonhuman primate model of inherited retinal disease. J Clin Invest 2019, 129:863-874

The first genetic model of eye disease in the primate, in this case a cone dystrophy. Effective pre-clinical development of vision restoration at the fovea could be accelerated if models of pan-retinal primate visual disfunction were more widely available.

- 69. Strazzeri JM, Hunter JJ, Masella BD, Yin L, Fischer WS, DiLoreto DA, Libby RT, Williams DR, Merigan WH: Focal damage to macaque photoreceptors produces persistent visual loss. *Exp Eye Res* 2014, **119**:88-96.
- Ivanova E, Hwang G-S, Pan Z-H, Troilo D: Evaluation of AAVmediated expression of Chop2-GFP in the marmoset retina. Invest Ophthalmol Vis Sci 2010, 51:5288-5296.

- 71. Mitchell JF, Leopold DA: **The marmoset monkey as a model for visual neuroscience**. *Marmoset Neurosci* 2015, **93**:20-46.
- 72. Sasaki E: Prospects for genetically modified non-human primate models, including the common marmoset. *Marmoset Neurosci* 2015, **93**:110-115.
- 73. Sato K, Sasaki E: Genetic engineering in nonhuman primates for human disease modeling. J Hum Genet 2018, 63:125-131.
- 74. Yin L, Masella B, Dalkara D, Zhang J, Flannery John G, Schaffer DV, Williams DR, Merigan WH: Imaging light responses of foveal ganglion cells in the living macaque eye. J Neurosci 2014, 34:6596.
- Pfäffle C, Hillmann D, Spahr H, Kutzner L, Kabuth B, Burhan S, Hilge F, Hüttmann G: Physiologic origin of intrinsic optical signals in human retina. Invest Ophthalmol Vis Sci 2018, 59:672.
- Ling T, Boyle KC, Goetz G, Zhou P, Quan Y, Alfonso FS, Huang TW, Palanker D: Full-field interferometric imaging of propagating action potentials. *Light Sci Appl* 2018, 7:107.
- 77. Yang HH, St-Pierre F: Genetically encoded voltage indicators: opportunities and challenges. J Neurosci 2016, 36:9977.
- Wang Q, Tuten WS, Lujan BJ, Holland J, Bernstein PS, Schwartz SD, Duncan JL, Roorda A: Adaptive optics microperimetry and OCT images show preserved function and recovery of cone visibility in macular telangiectasia type 2 retinal lesions. *Invest Ophthalmol Vis Sci* 2015, 56:778-786.
- 79. Russell S, Bennett J, Wellman JA, Chung DC, Yu Z-F, Tillman A, Wittes J, Pappas J, Elci O, McCague S et al.: Efficacy and safety of voretigene neparvovec (AAV2-hRPE65v2) in patients with RPE65-mediated inherited retinal dystrophy: a randomised, controlled, open-label, phase 3 trial. Lancet 2017, 390:849-860.
- Adeyemo O, Jeter PE, Rozanski C, Arnold E, Dalvin LA, Swenor B, Dagnelie G, PLoVR Study Group: Living with ultra-low vision: an inventory of self-reported visually guided activities by individuals with profound visual impairment Adeyemo et al. *Transl Vis Sci Technol* 2017, 6:10.
- Goldstein JE, Fenwick E, Finger RP, Gothwal V, Jackson ML, Lamoureux E, Rees G, Massof R: Calibrating the impact of vision impairment (IVI): creation of a sample-independent visual function measure for patient-centered outcomes research Goldstein et al. *Transl Vis Sci Technol* 2018, 7:38.
- Jeter PE, Rozanski C, Massof R, Adeyemo O, Dagnelie G, PLoVR Study Group: Development of the ultra-low vision visual functioning questionnaire (ULV-VFQ). Transl Vis Sci Technol 2017, 6:11.
- Petrillo J, Bressler NM, Lamoureux E, Ferreira A, Cano S: Development of a new Rasch-based scoring algorithm for the National Eye Institute Visual Functioning Questionnaire to improve its interpretability. *Health Qual Life Outcomes* 2017, 15:157.
- Markowitz M, Rankin M, Mongy M, Patino BE, Manusow J, Devenyi RG, Markowitz SN: Rehabilitation of lost functional vision with the Argus II retinal prosthesis. *Can J Ophthalmol* 2018, 53:14-22.
- 85. Komáromy AM, Alexander JJ, Rowlan JS, Garcia MM, Chiodo VA, Kaya A, Tanaka JC, Acland GM, Hauswirth WW, Aguirre GD: Gene therapy rescues cone function in congenital achromatopsia. *Hum Mol Genet* 2010, **19**:2581-2593.
- Carvalho LS, Xu J, Pearson RA, Smith AJ, Bainbridge JW, Morris LM, Fliesler SJ, Ding X-Q, Ali RR: Long-term and agedependent restoration of visual function in a mouse model of CNGB3-associated achromatopsia following gene therapy. *Hum Mol Genet* 2011, 20:3161-3175.
- Maguire AM, High KA, Auricchio A, Wright JF, Pierce EA, Testa F, Mingozzi F, Bennicelli JL, Ying G, Rossi S *et al.*: Age-dependent effects of RPE65 gene therapy for Leber's congenital amaurosis: a phase 1 dose-escalation trial. *Lancet* 2009, 374:1597-1605.